

Decay Products of the S_3 State of the Oxygen-Evolving Complex of Photosystem II at Cryogenic Temperatures. Pathways to the Formation of the $S = 7/2$ S_2 State Configuration[†]

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ABSTRACT: The water-oxidizing complex of photosystem II cycles through five oxidation states, denoted S_i ($i = 0-4$), during water oxidation to molecular oxygen, which appears at the (transient) S_4 state. The recent detection of bimodal EPR signals from the S_3 state [Matsukawa, T., Mino, H., Yoneda, D., Kawamori, A. (1999) *Biochemistry* 38, 4072–4077] has drawn significant attention to this critical state. An interesting property of the S_3 state is the sensitivity to near-IR (NIR) light excitation. Excitation of the S_3 state by near-IR light at cryogenic temperatures induces among other signals a derivative-shaped EPR signal at $g = 5$ [Ioannidis, N., and Petrouleas, V. (2000) *Biochemistry* 39, 5246–5254]. The signal bears unexpected similarities to a signal observed earlier in samples that had undergone multiple turnovers and subsequently had been stored at 77 K for a week or longer [Nugent, J. H. A., Turconi, S., and Evans, M. C. W. (1997) *Biochemistry* 36, 7086–7096]. Recently, both signals were assigned to an $S = 7/2$ configuration of the Mn cluster [Sanakis, Y., Ioannidis, N., Sioros, G., and Petrouleas, V. (2001) *J. Am. Chem. Soc.* 123, 10766–10767]. In the present study, we employ bimodal EPR spectroscopy to investigate the pathways of formation of this unusual state. The following observations are made: (i) The $g = 5$ signal evolves in apparent correlation with the diminution of the S_3 state signals during the slow (tens of hours to several days range) charge recombination of S_3 with Q_A^- at 77 K. The tyrosyl radical D^\bullet competes with S_3 for recombination with Q_A^- , the functional redox couple at cryogenic temperatures inferred to be D^\bullet/D^- . Transfer to -50°C and above results in the relaxation of the $g = 5$ to the multiline and $g = 4.1$ signals of the normal S_2 state. (ii) The transition of S_3 to the state responsible for the $g = 5$ signal can be reversed by visible light illumination directly at -30°C or by illumination at 4.2 K followed by brief (2 min) transfer to -50°C in the dark. The latter step is required in order to overcome an apparent thermal activation barrier (charge recombination appears to be faster than forward electron transfer at 4.2 K). (iii) The " $g = 5$ " state can be reached in a few tens of minutes at 4.2 K by near-IR light excitation of the S_3 state. This effect is attributed to the transfer of the positive hole from the Mn cluster to a radical (probably tyr Z), which recombines much faster than the Mn cluster with Q_A^- . (iv) The above properties strongly support the assignment of the configuration responsible for the $g = 5$ signal to a modified S_2 state, denoted S_2' . Evidence supporting the assignment of the S_2' to a proton-deficient S_2 configuration is provided by the observation that the spectrum of S_2 at pH 8.1 (obtained by illumination of the S_1 state at -30°C) contains a $g = 5$ contribution.

Photosystem II, PSII¹, catalyzes the light-induced oxidation of water to molecular oxygen. The site for this unique reaction is the so-called oxygen-evolving complex, OEC. OEC is the electron donor of PS II and cycles through five oxidation states $S_0 \dots S_4$ during light-induced sequential electron flow (for reviews, see 1–6).

EPR spectroscopy has played an important role in the study of the oxygen-evolving complex. EPR signals have now been detected from all S states, with the exception of the metastable S_4 state. The S_0 state is characterized by a $S =$

$1/2$ signal with weak hyperfine lines extending over a width of approximately 2800 G (7–9). The S_1 state is characterized by an integer spin signal at $g = 4.8$ with no resolved hyperfine structure (10, 11) or by an alternative multiline signal centered at $g = 12$ (12), both detected by parallel mode EPR. The S_2 state is characterized by the extensively studied

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¹ Abbreviations: PSII, Photosystem II; OEC, oxygen-evolving complex; WOC, water-oxidizing complex; S states $S_0 \dots S_4$ oxidation states of the water-oxidizing complex; tyr Z or Y_Z and tyr D or Y_D , the fast and slow tyrosine electron donors of PSII; signal II, the unperturbed EPR spectrum of either of the two tyrosines, a measure of oxidized tyr D under the present conditions; broad or split $g \sim 2$ signal, a broadened radical signal assigned to Tyr Z; Q_A and Q_B , the primary and secondary plastoquinone electron acceptors of PSII; BBY membranes, thylakoid membrane fragments enriched in PSII; MES, 2-[N-morpholineethanesulfonic acid]; chl, chlorophyll; EtOH, ethanol; MeOH, methanol; cw EPR, continuous wave electron paramagnetic resonance; NIR light, near-infrared light.

multiline signal ($S = 1/2$) at $g = 2$ (13) and the alternative $g = 4.1$ signal ($S \geq 3/2$) (14, 15). An unusual and interesting property of the S₂ state is the sensitivity to near-infrared light, NIR. The multiline signal can be converted to the $g = 4.1$ signal via an intermediate $S \geq 5/2$ state ($g = 10, 6$) by NIR illumination at temperatures greater than 60 K (16, 17).

Recently, low field EPR signals in both perpendicular and parallel mode EPR were detected from the S₃ state (18). The observations were subsequently confirmed, and interestingly a sensitivity of this state to NIR illumination was observed (19). These observations, which were made in plant material, were reproduced in a cyanobacterial preparation (20).

NIR-light excitation of the S₃ state produces, in addition to other signals that appear to be associated with excited-state configurations of S₃ (21), a derivative-shaped EPR signal at $g = 5$ (19). This signal bears unexpected similarities to a signal observed earlier by Nugent and co-workers in samples that had undergone multiple turnovers above S₁ and subsequently had been stored at 77 K for a week or longer (22). In a recent communication, the $g = 5$ signal was assigned to an $S = 7/2$ configuration of the S₂ state (23). The nearly isotropic $g = 5$ resonance originates from the "±3/2" doublet of the $S = 7/2$ manifold. A weaker resonance at around $g = 2.9$, whose position and intensity vary strongly with slight changes in the crystal-field parameters, results from the "±1/2" doublet.

In the present study, we correlate the $g = 5$ signal formation with the decay of the S₃-state signals and furthermore compare the two seemingly different mechanisms (long-term incubation at 77 K vs NIR excitation at liquid-helium temperatures), which, apparently, result in the same modified S₂ configuration. Evidence in support of the earlier proposal that this modified S₂ configuration is deprotonated (22), is provided by the observation that a $g = 5$ signal is produced directly by illumination of the S₁ state, at -30 °C and alkaline pH.

MATERIALS AND METHODS

PSII-enriched thylakoid membranes were isolated from market spinach by standard procedures (24, 25) with some modifications. Samples for EPR measurements were suspended in 0.4 M sucrose, 15 mM NaCl, 40 mM MES, pH 6.5, at 6–8 mg chl/mL and stored in liquid nitrogen until use.

Advancement to the S₃ state was achieved essentially as previously described (19). Samples (at 1 mg of chl/ml) were incubated with 2 mM ferricyanide at 4 °C, for 30 min, to oxidize the nonheme iron. The oxidant was subsequently removed by 3–4 wash steps in ferricyanide-free buffer by centrifugation at 35 000g. The final pellet was resuspended with oxidant-free buffer supplemented with 0.2 mM atrazine. The illumination procedure was as follows. BBY samples with preoxidized nonheme iron, initially kept at 4 °C in darkness, were rapidly transferred into a -30 °C precooled acetone bath under illumination, illuminated for 3 min, and finally placed in liquid nitrogen. For this purpose, white light from a 340 W projection lamp, filtered through a solution of CuSO₄, was used.

This procedure yielded approximately 50–60% S₃ and 50–40% S₂ state. The main contributions to the S₂ state presumably arose from centers in which the iron was not

initially oxidized and from a fraction of centers in which the S₃ state was reduced during illumination by Y_D. These centers should have Q_A singly reduced. In addition, a third contribution, representing a small fraction of S₂ centers with Q_A oxidized, presumably arose from the S₃/Q_A⁻ recombination during the illumination/freezing period.

Signal II, a measure of Y_D[•] in the present experiments, was monitored during all steps in the various experiments, and reference is made to its magnitude without showing the spectra, as they are trivial. Approximate normalization of signal II was done by assuming that the level of the signal at the end of the experiments and, following the full decay of S₃ and S₂ at about 15 °C, represented approximately 100% of the centers. This is a reasonable assumption considering that two electrons are transferred to the acceptor side—one to Q_A and one to Fe(III)—during formation of the S₃ state, but only Q_A⁻ is available for charge recombination (the oxidized iron is a stable electron acceptor). Thus, Y_D is the main alternative reductant of the excited S states (26). This must be added to the fact that a significant portion of this tyrosine is oxidized before the experiments (unless special precautions are taken) (27).

Excitation of the S₃ state by near-infrared light (NIR) was performed at 4.2 K. The projection lamp was filtered in this case by a 3 mm RG715 SCHOTT filter immersed in water. Illumination with white light was done in the same manner, but the projector light was filtered by a 3 mm BG-39 SCHOTT filter immersed in a CuSO₄ solution. The duration of illuminations was, unless otherwise stated, 3 min intermittently in 15 s intervals.

At the end of experiments, samples were dark adapted for typically 60 min at 4 °C, or 30 min at 15 °C, to relax to the S₁ state. The resulting EPR spectrum was used as a reference and was subtracted from all other spectra in order to eliminate background contributions due to oxidized cyt *b*₅₅₉ and impurities. Following the dark adaptation period, samples were illuminated again at -30 °C in order to obtain the maximum S₂ state population.

The S₁-to-S₂ advancement at alkaline pH was performed as follows. PS II enriched membranes were centrifuged at 35 000g and resuspended twice in a buffer containing 0.4 M sucrose, 15 mM NaCl, 1 mM MES, pH 6.5. In the second resuspension step, the buffer was supplemented with 0.2 mM atrazine. Samples were illuminated at -30 °C for 2 min, and the S₂ state (pH 6.5) control spectra were recorded. The samples were subsequently allowed to decay to the S₁ state by dark adaptation at 15 °C, and the pH was raised to approximately 8 (measured at the end of the experiments) by the addition of 40 mM final concentration Tricine from a stock solution of 0.5 M, pH 8.5. After mixing (<30 s), the samples were either frozen immediately or allowed to equilibrate for up to 30 min at 15 °C (no significant differences were observed among the samples), and the alkaline S₁ spectra were recorded. The S₂ state spectrum was obtained subsequently after illumination at -30 °C.

EPR measurements were performed with a Bruker ER-200D-SRC spectrometer interfaced to a personal computer and equipped with an Oxford ESR 900 cryostat, an Anritsu MF76A frequency counter, and a Bruker 035M NMR gaussmeter. Where not specified, the perpendicular 4102ST cavity was used, and the microwave frequency was 9.41 GHz with 4 mm and 9.39 GHz with 5 mm tubes. The spectra

obtained with the standard cavity are the average of 2–4 scans. Tubes of 5 mm were exclusively used with the dual mode cavity, 4116 DM, and 4 or 10 accumulations were done in the perpendicular or the parallel mode, respectively. The microwave frequency was 9.60 GHz in the perpendicular and 9.31 GHz in the parallel mode.

RESULTS AND DISCUSSION

Decay of the S_3 State at 77 K. Nugent et al. (22) reported a signal at about $g = 5$ in samples that had undergone multiple turnovers above the S_1 state and subsequently had been stored at 77 K for more than 1 week. The signal was assigned to a modified S_2 state, which accumulated during the decay of the S_3 state at 77 K; however, alternative possibilities, e.g., assignment of the signal to a modified form of the oxidized nonheme iron, could not be excluded. In view of the recent detection of EPR signals from the S_3 state (19), we have examined the time course of its decay and the concomitant evolution of other signals at 77 K. This is depicted in Figure 1. A sample with the nonheme iron preoxidized was illuminated as described in the Materials and methods section and thus advanced to the S_3 state, trace a. Then, the sample was incubated at 77 K in the dark, and spectra were collected after 17 h (trace b), 36 h (trace c), and 172 h (trace d). The following observations can be made. During incubation at 77 K, the signals assigned to the S_3 state ($g = 10$), the S_2 state (multiline and $g = 4.1$), and the Q_A^- -Fe(II) complex decrease gradually; in addition, signal II (a measure of oxidized Y_D under the present conditions, see also Materials and Methods) decreases (see later in this section) due to recombination with Q_A^- (28, 22).

The above changes are accompanied by the gradual formation of a signal at $g = 5$ (apparently the same signal reported by Nugent et al., 22). The intensity of the signal becomes maximal after incubation of the S_3 state at 77 K for about 1 week (Figure 1d). The signal is more pronounced at 4.2 K but at 11 K becomes significantly weaker. Small contributions from the $g = 4.1$ signal (S_2 state) distort somewhat its shape, particularly at 11 K (at 4.2 K the S_2 multiline and $g = 4.1$ signals are relatively smaller due to microwave power saturation). Incubation of samples at temperatures between 77 and 180 K, following incubation at 77 K for a few days, results in no significant changes except for a slight narrowing of the $g = 5$ signal (separate experiment not shown).

Warming of the sample in Figure 1d to -80°C for 2 min causes a 30% decrease of the $g = 5$ signal and subsequent warming to -50°C for another 2 min, Figure 1e, results in an almost complete loss of the $g = 5$ signal and a decrease of the $g = 10$ signal. A significant enhancement of the S_2 multiline and $g = 4.1$ signals,² as well as an increase in the signal II size (see later in this section) accompanies the last changes. Signal II reaches its maximum size (100%, see Materials and Methods) following a subsequent 30 min dark adaptation at 15°C .

² We confirmed in a separate experiment, using the standard cavity, the observation of Nugent et al. (22) that the restored S_2 multiline form at -80°C is slightly different than the form obtained by illumination of S_1 samples at the same temperature. Changes in the $g = 4.1$ signal could not be positively detected due to interference from the $g = 5$ signal.

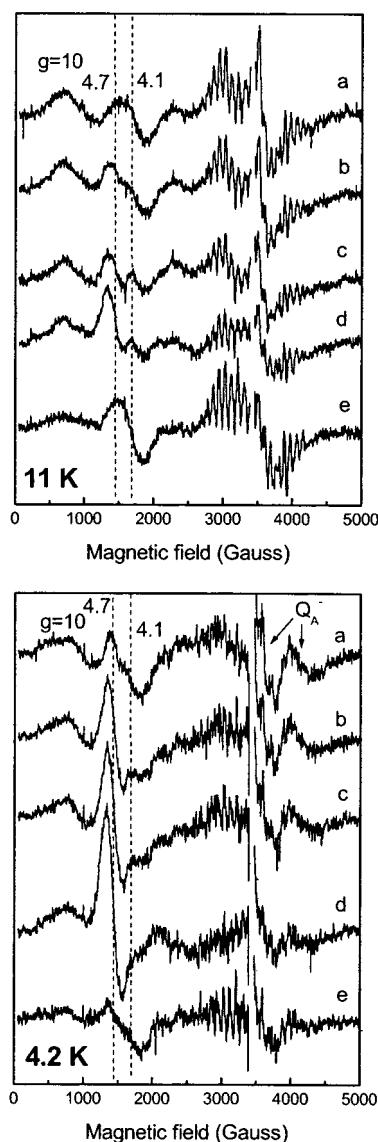


FIGURE 1: Time-dependent evolution of the $g = 5$ signal and diminution of the $g = 10$ signal during incubation of the S_3 state at 77 K. Spectrum a represents the S_3 state as prepared. Spectra b–d were recorded after 17, 36, and 172 h, respectively, of incubation time of the sample in trace a at 77 K. Finally, the sample was transferred to -80°C for 2 min and subsequently to -50°C for 2 min, spectrum e. All spectra are difference traces obtained after subtraction of the final dark-adapted spectrum as described in the Materials and Methods section. EPR conditions: temperature as indicated, microwave frequency 9.59 GHz, microwave power 35 mW, modulation amplitude 10 G, modulation frequency 100 kHz.

The S_3 decay has also been examined by parallel-mode EPR. Figure 2 shows the initial level of the S_3 signal (peak at $g = 17$) and the level of the signal after 82 h of incubation at 77 K. A significant decay of the S_3 signal is observed in agreement with the perpendicular-mode experiment of Figure 1.

The results of three experiments similar to those of Figures 1 and 2 are summarized in the diagrams of Figure 3, where the amplitude of the signals is plotted against incubation time at 77 K. The advancement to the S_3 state is taken as time zero, while the amplitudes of the various signals are normalized to the percentage of PSII centers they represent (see Materials and Methods). The calculation of the S_2 state fraction was based on the amplitude of the multiline signal,

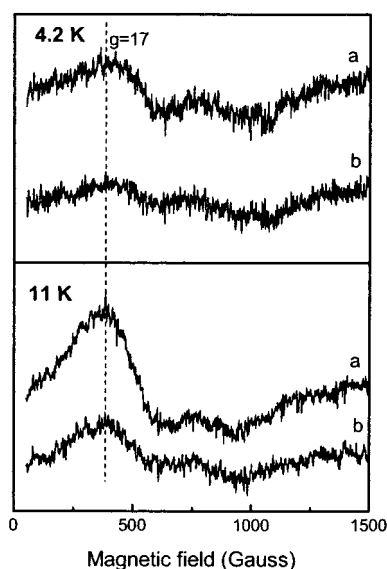


FIGURE 2: Decay of the S₃-state parallel-mode spectrum during prolonged storage at 77 K. Spectra, at the indicated temperatures, were recorded (a) immediately after the formation of the S₃ state and (b) after a subsequent 82 h incubation at 77 K. EPR conditions: microwave frequency 9.30 GHz, microwave power 8.2 mW at 4.2 K and 129 mW at 11 K, modulation amplitude 10 G, modulation frequency 100 kHz.

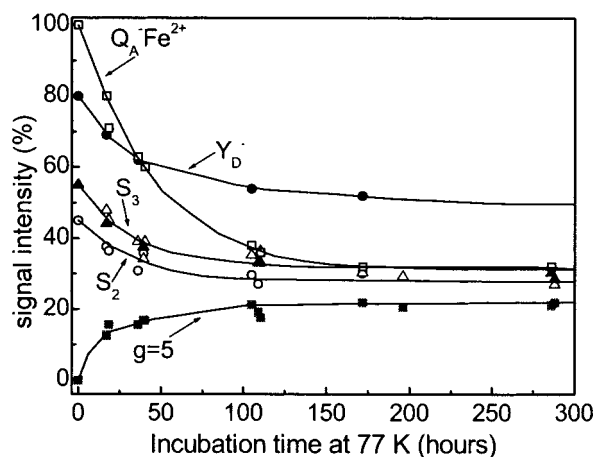


FIGURE 3: Time course of changes observed in the amplitude of various signals affected by the prolonged incubation of the S₃ state at 77 K. Filled squares, the $g = 5$ signal (modified S₂ state); open circles, the multiline signal (normal S₂ state); open triangles, the $g = 10$ signal (S₃ state, perpendicular mode); filled triangles, the $g = 17$ signal (S₃ state, parallel mode); filled circles, signal II slow (Y_D[•]); open squares, the Q_A⁻ semiquinone radical signal. Data points from three different experiments are pooled. The normalization of the signal amplitudes was done as described in the Materials and Methods section.

since the $g = 4.1$ signal is distorted by contributions from the $g = 5$ signal. Different symbols are used for the perpendicular-mode ($g = 10$, open triangles) and parallel-mode ($g = 17$, filled triangles) signals of the S₃ state, but it is reassuring to note that the decay traces coincide within experimental error. The following general remarks can be made comparing the intensity losses of the various signals. The Q_A⁻Fe²⁺ signal decreases by about 70%, presumably due to charge recombination with Y_D[•] (22) in 28% of the centers and S₂ in 15% of the centers (these are centers where S₂ and Q_A⁻ coexist, see Materials and Methods). The remaining fraction is consistently accounted for by the 25%

decay of S₃. These assignments are supported by the considerations below.

The $g = 5$ signal in Figure 1 is undoubtedly the same signal that has been reported by Nugent et al. (22). In the present case, atrazine restricts sample advancement beyond the S₃ state. In the absence of atrazine, both the size of the initial S₃ signal and the $g = 5$ signal that developed during 15 days of storage at 77 K were smaller compared to the control sample that contained the inhibitor atrazine (spectra not shown). On the other hand, we have confirmed the observations of Nugent et al. (22) that, samples poised at the S₂ state did not develop the $g = 5$ signal on storage at 77 K. In Figure 3a, correlation exists between the decay of the S₃ signals and the evolution of the $g = 5$ signal. The S₃-state EPR signals are indicative of an even number of electrons in this state. The $g = 5$ signal on the other hand has been assigned to a half-integer spin, $S = 7/2$ (23), indicative of a configuration with an odd number of electrons. The most reasonable interpretation of the above results is that the evolution of the $g = 5$ signal is due to the reduction of the S₃ state at 77 K by one electron (due to charge recombination with Q_A⁻ as was assumed above). At elevated temperatures (-50 °C), the $g = 5$ signal is converted to the normal S₂ multiline and $g = 4.1$ signals (Figure 1e, see also ref 22). Hence, the $g = 5$ signal is assigned to a modified S₂ configuration (we will subsequently call it S₂[']), as was originally suggested (22). This interpretation is strongly supported by the fact that the decay of the S₃ state is reversed by white-light illumination (see below).

It is notable that the S₃ state does not decay fully at cryogenic temperatures, Figure 3. A nondecaying fraction of S₃ is observed even after very long incubation at 77 K (see also 21). The results of Figure 3 indicate that Y_D[•] is reduced concomitantly to the decay of the S₃ state. Presumably, Y_D[•] and the S₃ state, which coexist in a significant fraction of centers, compete for charge recombination with Q_A⁻, although direct redox exchange between Y_D[•] and S₃ is also expected. It is reasonable to assume that during prolonged incubation at 77 K redox equilibrium is established between the Mn cluster and Y_D. There is general consensus that at ambient temperatures, Y_D is neutral in both the oxidized and the reduced form (for a recent review, see ref 29). At cryogenic temperatures, exchange of a proton between this tyrosine and its surroundings is probably prohibited. This is suggested by the fact that Y_D, when frozen to cryogenic temperatures in the neutral-reduced state, does not function as a low temperature donor. However, when frozen in the oxidized state, Y_D[•], is slowly reduced at 77 K by charge recombination with Q_A⁻ to the nonprotonated form, Y_D⁻, and then becomes a very efficient electron donor at cryogenic temperatures. Illumination, in this case, with visible light at temperatures as low as 4.2 K, results readily in charge separation between Y_D⁻ and Q_A (22, 21). The above observations can be rationalized if we assume that the functioning redox couple at 77 K is Y_D[•]/Y_D⁻. It is possible that the two couples, Y_D[•]/Y_D⁻ and S₃/S₂ ($g = 5$) have similar redox potentials.

A brief comment should be also made about the annealing step to -50 °C (Figure 1e). Recombination with Q_A⁻ is slower than the 2 min interval at this temperature (see next section), but Tyr D is expected to act as a rapid reductant of S₃ and S₂ in those centers where the reduced tyrosine (formed

mainly during the prolonged incubation at 77 K, Figure 3) coexists with either of the two states (28, 30, 31). As the $g = 5$ signal converts to the S_2 multiline and $g = 4.1$ forms, the net effect of the 2 min incubation at -50°C , Figure 1e, is a decrease of the S_3 signals, and an enhancement of the S_2 signals. In addition, the oxidized fraction of tyr D is approximately restored to its level in Figure 1a. It should be noted that the decay of S_3 and S_2 at -50°C is slow when Y_D is oxidized (see below). It has been also observed that the conversion of the $g = 5$ to the multiline and $g = 4.1$ signals is independent of the redox state of Y_D .

Decay of the S_3 State at Elevated Temperatures. It was noted above that no new signals develop during the incubation of the $g = 5$ containing samples at temperatures between 77 and 180 K, while at -50°C the $g = 5$ signal relaxes within a few minutes to the S_2 multiline and $g = 4.1$ signals (see, however, footnote 2). On the other hand, we could not find a convenient temperature higher than 77 K that accelerated the reduction rate of the S_3 state to the $g = 5$ configuration. Most likely the decay of the $g = 5$ signal at elevated temperatures is faster than its production. At ca. 170 K, the $t_{1/2}$ for the decay of the S_3 state signal at $g = 10$ is longer than 8 h, while the $g = 5$ decays with an approximately half-time of a few hours. At -50°C , the approximate half-time for the decay of S_3 , due to recombination with Q_A^- , (samples with Y_D fully oxidized) is 30 min, while the $g = 5$ signal decays fully within 2 min. The decay products of the S_3 state at -50°C are the S_2 multiline and $g = 4.1$ signals, at relative ratios approximately comparable to the native S_2 state. A typical sequence of events describing the decay of the S_3 state at elevated temperatures is exemplified by an experiment performed at -30°C (Figure 4).

Following the initial population of the S_3 state (uppermost spectrum), spectra were collected after various time periods of incubation at -30°C . At the end of the incubation period at -30°C and after 30 min of additional dark adaptation at 15°C —this allowed for the complete decay to the S_1 state—the sample was illuminated at 200 K, and the spectrum of the S_2 state was recorded as a reference. By comparison with the latter spectrum and assuming that the S_3 and S_2 are the main states occupied, the starting populations of the S_3 and S_2 states in Figure 4 are estimated at approximately 60% and 40% respectively. During incubation at -30°C , the S_3 state signal at $g = 10$ declines continuously, but no clear buildup of the S_2 multiline or $g = 4.1$ signals is evident. It is clear, however, that the decline of the S_2 signals is significantly slower. The data are difficult to interpret quantitatively. A simple qualitative explanation would be that the initial S_3 and S_2 fractions recombine with Q_A^- with similar rate constants, while the fraction of S_2 resulting from the decay of S_3 is reduced more slowly by alternative reductants.

We have also examined a simpler case for comparison. In the presence of 5% (v/v) methanol, the S_2 state is represented by the multiline signal only (32). It was also noted in our earlier study (19) that no S_3 signals could be detected in the presence of methanol—occupation of this state was inferred by the diminished amplitude of the multiline signal—and no sensitivity to NIR light was observed. Thus, in an experiment similar to that of Figure 4, we examined the decay of the S_3 state in the presence of methanol (not shown). In this case, we did find a clear transient increase

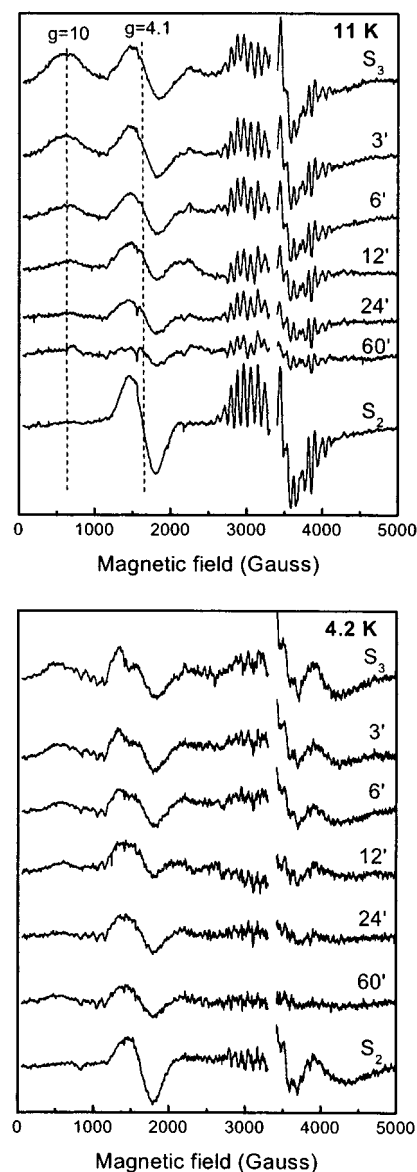


FIGURE 4: Decay of the S_3 state at -30°C . Spectra labeled S_3 were recorded immediately after excitation to the S_3 state, and subsequent spectra were collected after incubation at -30°C for the indicated time periods. The bottom spectrum represents the maximal intensity of the S_2 state. Difference spectra were obtained by subtraction of the dark-adapted spectrum obtained at the end of the measurements (see Materials and Methods). EPR conditions: temperature as indicated, microwave frequency 9.39 GHz, microwave power 35 mW, modulation amplitude 25 G, modulation frequency 100 kHz.

of the S_2 state multiline, which declined at longer times. A gradual decay of Q_A^- was observed in parallel.

The data on the $S_3Q_A^-$ recombination rate are scarce (33, 34), but it is usually assumed that it is similar to that of the $S_2Q_A^-$ recombination. This is approximately 1 s at room temperature (33, 34) and slows down to a $t_{1/2}$ of ca. 5 min at -25°C (35, 36). This estimate is supported by the decay rate of the $g = 10$ (S_3) and the $Q_A^-Fe^{2+}$ signals in Figure 4. In this experiment, the starting population of the S_3 state is higher than that of the S_2 state. If the decay process of the $g = 10$ signal to the multiline and $g = 4.1$ signals is first-order, one will expect, under these conditions, a transient built up of the S_2 signals, which is indeed observed in the presence of methanol (not shown). The absence of a transient

increase of the S₂ signals in Figure 4 may indicate a slow conversion to the conformations that produce the S₂ signals (see also ref 2). Alternatively, the S₂Q_A⁻ recombination rate may be somewhat faster than that of the S₃Q_A⁻ in the untreated sample and slower in the MeOH treated one.

Recovery of the S₃ State, Following Its Decay, by Visible Light Illumination. The main states generated by the procedure used to form the S₃ state are S₃Q_A⁻ and S₂Q_A⁻ (see Materials and Methods). Charge recombination at -30 °C or lower (as described in the previous section) should produce the S₂Q_A and S₁Q_A states, respectively. Provided that the incubation at the elevated temperatures is not prolonged (that would lead to the slow reduction of S₂Q_A to S₁Q_A), illumination at -30 °C should induce charge separation and restore the initial states. This has been readily confirmed with samples advanced to the S₃ state and subsequently incubated at -50 °C long enough for charge recombination to occur. Since the *g* = 5 configuration is assigned to a modified S₂, S₂', a similar behavior should be exhibited by samples where the S₃ decayed after prolonged incubation at 77 K. This is demonstrated by the following two experiments.

In an experiment similar to Figure 1, a sample poised in the S₃ state, Figure 5a, was incubated at 77 K for 11 days until the *g* = 10 signal decreased considerably and a pronounced *g* = 5 signal developed, Figure 5b. The sample was subsequently incubated at -50 °C for 2 min, Figure 5c (a treatment similar to Figure 1e). Notable is the significant decrease of the Q_A⁻Fe²⁺ signal in traces b and c of Figure 5, indicating that charge recombination has occurred in the largest fraction of centers. As discussed in the first section, the initial level of tyr D• is approximately restored at -50 °C; therefore, recombination must have occurred at the expense of the S₃ and S₂ states at this final stage. Assuming roughly equal initial populations of S₂Q_A⁻ and S₃Q_A⁻, the spectra of Figure 5c consist mainly of approximately equal populations of S₁Q_A and S₂Q_A (in addition to small portions of not recombined S₂Q_A⁻ and S₃Q_A⁻). Illumination at -30 °C, Figure 5d, clearly restores the starting population of states.

In the discussion related to Figures 1–3, it was concluded that the one-electron reduction of the S₃ state at 77 K gives rise to the S₂' state (characterized by the *g* = 5 signal). It is reasonable to expect that white-light illumination of the latter state at cryogenic temperatures will result in the formation of the initial S₃ state. The fact that this is possible has been implied by the experiments of Nugent et al. (22). These authors used samples which developed the *g* = 5 signal and noticed the induction of charge separation upon illumination at liquid helium temperatures. This charge separated couple recombined when stored at liquid helium temperatures, whereas at temperatures above 77 K, it was stabilized. Accordingly, the advancement of the *g* = 5 configuration of the Mn cluster to a higher oxidation state appeared to have an activation barrier below liquid nitrogen temperatures. Figure 6 repeats in a somewhat modified form the original experiment of Nugent et al. with the added advantage of knowing now the signature of the S₃ state. The experiment was followed by both perpendicular and parallel - mode EPR. Spectrum a shows the S₃ state as prepared. Spectrum b was recorded after several weeks of incubation at 77 K. It can be seen that the intensity of the S₃ signals, i.e., at *g* = 10 in perpendicular mode and *g* = 17 in parallel mode, is

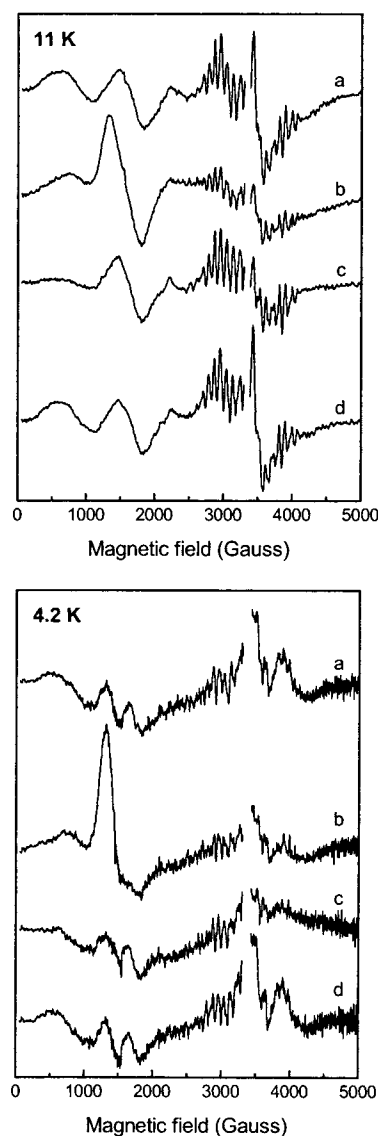


FIGURE 5: Recovery of the S₃ state, following its decay at 77 K, by visible light illumination at -30 °C. Spectra were recorded at 11 and 4.2 K after the following successive treatments of the same sample. (a) The S₃ state as prepared. (b) Incubation at 77 K for 11 days. (c) Warming to -50 °C for 2 min. (d) Illumination at -30 °C for 3 min. Difference spectra were obtained by subtraction of the dark adapted spectrum as described in the Materials and Methods. EPR settings are the same as those in Figure 4.

considerably diminished while a strong signal at *g* = 5 has appeared. Evident also is the great diminution of the Q_A⁻Fe(II) signal. Visible-light illumination at 4.2 K of the sample, followed immediately by transfer to -50 °C, spectrum c (see ref 21 for the intermediates of this experiment), restores the Q_A⁻Fe(II) signal to a high extent and causes the formation of a substantial amount of the S₃ state, as evidenced by the reformation of the *g* = 10 and 17 signals.

Clearly, the efficiency of the experiment of Figure 6 cannot be as high as of the one in Figure 5. In Figure 5, reduction of S₃ by tyr D at -50 °C increased the fraction that had decayed to S₂ by the previous storage at 77 K. Furthermore, the illumination at -30 °C is far more efficient in producing charge separation than the illumination at liquid-helium temperatures. Nevertheless, recovery of the S₃ state by the unusual protocol of Figure 6 is clearly demonstrated.

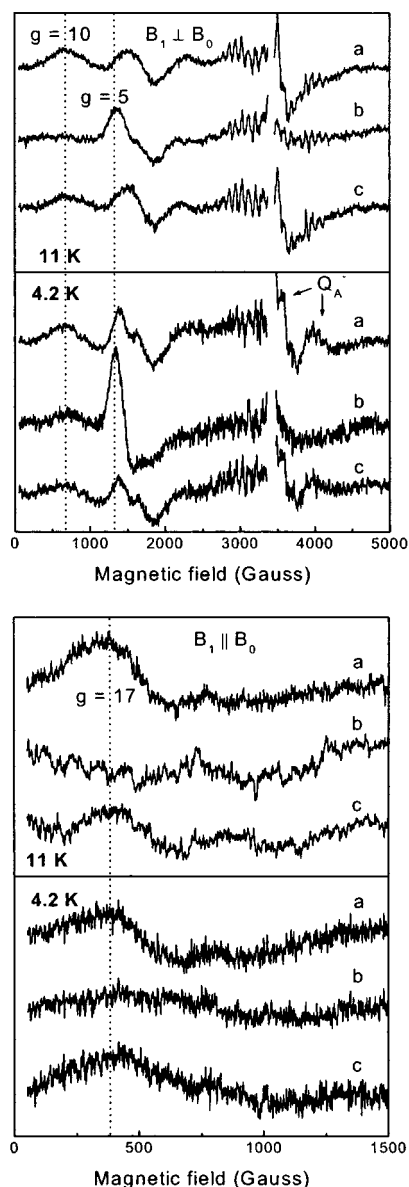


FIGURE 6: Recovery of the S_3 state, following its decay at 77 K, by visible light illumination at 4.2 K. Spectra at perpendicular and parallel modes were recorded after the following successive treatments of the same sample. (a) The S_3 state as prepared. (b) Incubation at 77 K for several weeks. (c) Illumination at 4.2 K for 3 min, immediately followed by transfer to -50°C for 2 min. Difference spectra were obtained by subtraction of the dark adapted spectrum as described in the Materials and Methods section. Experimental conditions were the same as those in Figure 1 for the perpendicular-mode panels and as those in Figure 2 for the parallel-mode panels. Recording temperatures as indicated.

NIR Excitation of the S_3 State Accelerates Charge Recombination. The NIR excitation at 50 K of the S_3 state in (19) produced, in addition to other signals that appear to be associated with excited-state configurations of S_3 (21), a $g = 5$ signal, which shows notable similarities to the signal in Figure 1 produced slowly during storage of the S_3 state at 77 K. It was noted during the present studies that the changes induced by the NIR-light excitation of the S_3 state at about 50 K can be resolved better if the NIR-light excitation is performed at 4.2 K. A systematic study is presented in the accompanying paper (21). Here we focus on the formation of the $g = 5$ signal.

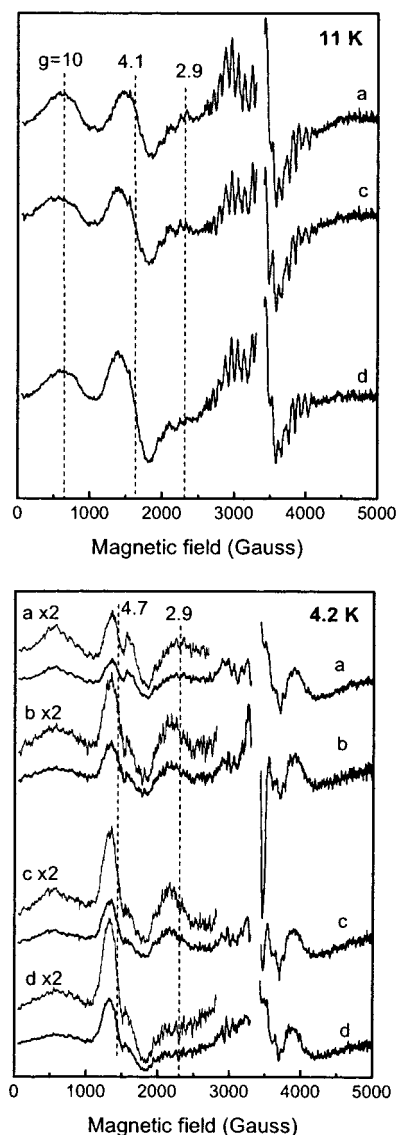


FIGURE 7: Induction of the $g = 5$ signal by excitation of the S_3 state with NIR light. Spectra were recorded after the following successive treatments of the same sample: (a) the S_3 state as prepared, (b) immediately after NIR excitation at 4.2 K, (c) 100 min incubation at 4.2 K and 30 min incubation at 11 K, and (d) 15 min incubation at 77 K. Difference spectra were obtained after subtraction of the dark adapted spectrum as described in the Materials and Methods section. EPR conditions: temperature as indicated in the panels, microwave frequency 9.39 GHz, microwave power 35 mW, modulation amplitude 25 G, modulation frequency 100 kHz.

Figure 7a shows the spectrum of the S_3 state as prepared. The sample was subsequently excited by NIR light at 4.2 K. This resulted in the parallel formation of the broadened $g \sim 2$ signal and the $g = 5$ signal with the 2.9 component (19, 23), Figure 7b. The spectrum recorded following a dark adaptation period at 4.2 and 11 K, which allowed for an approximately 70% decay of the $g = 2$ signal, is shown in Figure 7c. A small increase/sharpening of the $g = 5$ signal can be observed. Finally, the sample in c was transferred for 15 min at 77 K, and the new spectrum was recorded, Figure 7d. This resulted in the elimination of the $g = 2.9$ signal and a narrowing and increase of the $g = 5$ signal. These changes are more clearly seen in the spectra at 4.2 K. At 11 K, the $g = 5$ signal is obscured by the $g = 4.1$ signal

from the portion of centers in the S₂ state. The latter signal is partially saturated and thus makes a smaller contribution at 4.2 K. It should be noted that the NIR illumination had no effect on the amplitude of Y_D[•].

In a recent communication (23), the $g = 5/2.9$ signals induced by NIR light, as well as the $g = 5$ signal that results from the slow decay of the S₃ state at 77 K, were assigned to the same $S = 7/2$ configuration of the Mn cluster. Small changes in the crystal field parameters (reflecting structural changes), induced by the different treatments (NIR excitation vs incubation at 77 K), cause different relative amplitudes of the $g = 5$ and 2.9 features. While a more thorough analysis (see also 37), including the orientation dependence of the signals is currently in progress, a number of qualitative remarks can be made on the spectra.

The nearly-isotropic " $g = 5$ only" spectrum, obtained after the prolonged incubation of the S₃ state at 77 K, Figures 1d and 6b, appears to represent the most relaxed configuration of the $S = 7/2$ state. The $g = 5/2.9$ spectrum obtained by NIR-excitation appears to represent a transient configuration. The gradual conversion to the nearly isotropic form, e.g., spectra c and d of Figure 7, results in a sharpening and increase of the $g = 5$ component, but as can be verified by theory (23, 37), the number of centers contributing to the spectra remains the same. This argues against the possibility that the increase in the $g = 5$ signal in spectrum c relative to spectrum b in Figure 7 is at the expense of the $g = 2$ radical. Spectrum b in this figure was recorded immediately after the NIR excitation, while spectrum c was recorded after the decay of the largest fraction of the radical. A close inspection of several experiments with varying NIR-excitation periods indicated that the $g = 5$ signal forms concomitantly to the $g = 2$ broad radical. What may accordingly affect the apparent intensity of the $g = 5$ signal is a change in the crystal field parameters in the presence and the absence of the radical species. It is possible, e.g., that during formation of the radical, proton shifts and Coulombic interactions cause small changes to the bond lengths/angles in the Mn cluster, resulting in a small and negative value of the D parameter of the empirical spin Hamiltonian (23). After decay of the radical, a slow rearrangement of the cluster occurs, which results in positive D values. An alternative interpretation could be that the magnetic interaction of the radical with the Mn cluster, which results in the broadening of the radical spectrum, broadens also the $g = 5/2.9$ spectrum. Preliminary theoretical simulations indicated, however, that a magnetic coupling that would produce a significant broadening of the radical signal would induce a nonobservable broadening in the $g = 5/2.9$ spectrum.

The $S = 7/2$ configuration is assigned to a modified S₂ state (denoted S₂[']), as was argued earlier in this paper. The rapid attainment by the NIR-light-excitation of the same modified S₂ configuration of the Mn cluster that results from the slow reduction of the S₃ state at 77 K is astonishing. It appears that the NIR-light excitation of the freshly prepared S₃ state accelerates charge recombination in some of the centers. Conceivably, this is possible if we assume that the NIR excitation causes the transfer of the positive hole from the Mn cluster to a nearby residue forming the broadened $g = 2$ radical. The radical species (tyr Z[•], as argued in the following paper, 21) in turn recombines rapidly with Q_A⁻ to yield an excited S₂ state configuration, S₂['], identical to

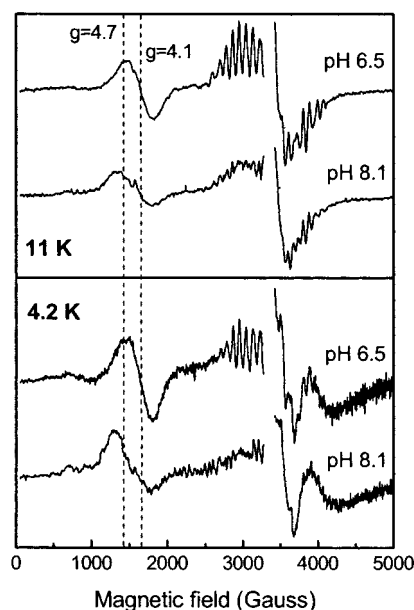


FIGURE 8: S₂ state spectra at alkaline pH contain a $g = 5$ contribution. The same sample was incubated first at pH 6.5 and subsequently at pH 8.1 (see Materials and Methods). The S₂ state was produced by illumination of the S₁ state at -30°C in the presence of the atrazine inhibitor. Difference spectra were obtained by subtraction of the respective S₁ traces at each pH. EPR settings were the same as those in Figure 4. Recording temperatures as indicated.

the one that evolves during the very slow recombination of S₃ with Q_A⁻ at 77 K. One would accordingly expect a small decrease in the intensity of the Q_A⁻Fe²⁺ signal accompanying the decay of the radical signal. As the inherently weak Q_A⁻Fe²⁺ signal in Figure 7a represents 100% of the centers while the radical species represents less than 10% of the PSII centers (19), such a decrease would be difficult to detect. Nevertheless, a small decrease can be discerned in the spectra c and d of Figure 7. Alternatively, in preliminary experiments using the exogenous electron acceptor phenyl-p-benzoquinone (38), the NIR-induced radical species was very stable over a period of 4 h at 11 K in the absence of Q_A⁻ (data not shown).

S₂ State Spectra at Alkaline pH Contain a $g = 5$ Contribution. It was shown in the preceding experiments that incubation at 77 K or NIR excitation of the S₃ state at 4.2 K invariably result in the decay of S₃ to S₂[']. The latter is a modified S₂ configuration, most likely deficient by one proton, as originally suggested (22). The relaxation of S₂['] to the normal S₂ configuration at -50°C and higher temperatures is accordingly attributed to protonation of the Mn cluster (not excluding parallel conformational changes).

The above interpretation is supported by experiments, which show that production of the S₂ state by the straightforward illumination of S₁ at -30°C at alkaline pH is accompanied by the formation of a $g = 5$ signal. An experiment performed as described in the Materials and Methods section is depicted in Figure 8. This compares the S₂ state spectra of the same sample at pH 6.5 and 8.1. The intensity of the S₂ state signals (multiline and $g = 4.1$) is greatly reduced on going from pH 6.5 to 8.1. Notable is, however, the formation of a $g = 5$ signal at pH 8.1.

The decrease of the multiline signal at high pH is compatible with the systematic observations of Geijer et al.

(39), but as these authors used 5% methanol, which is known to enhance the S_2 multiline signal at the expense of alternative signals, no comparison can be made with the changes in the low-field EPR signals of Figure 8. Geijer et al. (39) attributed the diminution of the multiline signal at high pH to protonation/deprotonation of ligands to the Mn (other possibilities were considered as well). This agrees with the present observations. Actually, the fact that the same $g = 5$ conformation appears to be trapped during the decay of S_3 at 77 K (or even at 4.2 K by the NIR excitation) as well as during the light-induced advancement of the S_1 to S_2 state at alkaline pH suggests deprotonation of the same Mn ligand in both cases. Clearly, the high-pH effects deserve a more systematic study before final conclusions can be made.

The NIR excitation of the S_3 state at liquid-helium temperatures, produces excited- S_3 -state transients in addition to the $g = 5/2.9$ signal (19). These, as well as the products of the light excitation of the S_2' state, are studied in the companion paper (21). The overall phenomenology is examined in the context of a molecular model, which provides useful insights into the interrelation of the Mn cluster and tyr Z.

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REFERENCES

1. Britt, R. D. (1996) *Advances in Photosynthesis: Vol. 4 Oxygenic Photosynthesis: The Light Reactions* (Ort, R. D., and Yocum, C. F., Eds) pp 137–164, Kluwer Academic Publishers, Dordrecht, The Netherlands.
2. Diner, B. A. and Babcock, G. T. (1996) *Advances in Photosynthesis: Vol. 4 Oxygenic Photosynthesis: The Light Reactions* (Ort, R. D., and Yocum, C. F., Eds) pp 213–247, Kluwer Academic Publishers, Dordrecht, The Netherlands.
3. Rutherford, A. W. (1989) *Trends Biol. Sci.* 14, 227–232.
4. Debus, R. J. (1992) *Biochim. Biophys. Acta* 1102, 269–352.
5. Bricker, T. M.; Ghanotakis, D. F. (1996) *Advances in Photosynthesis: Vol. 4 Oxygenic Photosynthesis: The Light Reactions* (Ort, R. D., and Yocum, C. F., Eds) pp 113–136, Kluwer Academic Publishers, Dordrecht, The Netherlands.
6. Yachandra, V. K., Sauer, K., and Klein, M. P. (1996) *Chem. Rev.* 96, 2927–2950.
7. Messinger, J., Nugent, J. H. A., and Evans, M. C. W. (1997) *Biochemistry* 36, 11055–11060.
8. Åhring, K. A., Peterson, S., and Styring, S. (1997) *Biochemistry* 36, 13148–13152.
9. Messinger, J., Robblee, J. H., Wa On Yu, Sauer, K., Yachandra, V. K., and Klein, M. P. (1997) *J. Am. Chem. Soc.* 119, 11349–11350.
10. Dexheimer, S. L., and Klein, M. P. (1992) *J. Am. Chem. Soc.* 114, 2821–2826.
11. Yamauchi, T., Mino, H., Matsukawa, T., Kawamori, A., and Ono, T. (1997) *Biochemistry* 36, 7520–7526.
12. Campbell, K. A., Gregor, W., Pham, D. P., Peloquin, J. M., Debus, R. J., and Britt, R. D. (1998) *Biochemistry* 37, 5039–5045.
13. Dismukes, G. C., and Siderer, Y. (1981) *Proc. Natl. Acad. Sci. U.S.A.* 78, 274–278.
14. Casey, J. L., and Sauer, K. (1984) *Biochim. Biophys. Acta* 767, 21–28.
15. Zimmermann, J. L., and Rutherford, A. W. (1984) *Biochim. Biophys. Acta* 767, 160–167.
16. Boussac, A., Girerd, J.-J., and Rutherford, A. W. (1996) *Biochemistry* 35, 6984–6989.
17. Boussac, A., Un, S., Horner, O., and Rutherford, A. W. (1998) *Biochemistry* 37, 4001–4007.
18. Matsukawa, T., Mino, H., Yoneda, D., Kawamori, A. (1999) *Biochemistry* 38, 4072–4077.
19. Ioannidis, N., and Petrouleas, V. (2000) *Biochemistry* 39, 5246–5254.
20. Boussac, A., Sugiura, M., Inoue, Y., and Rutherford, A. W. (2000) *Biochemistry* 39, 13788–13799.
21. Ioannidis, N., Nugent, J. H. A., and Petrouleas, V. (2002) *Biochemistry* 41, 9589–9600.
22. Nugent, J. H. A., Turconi, S., and Evans, M. C. W. (1997) *Biochemistry* 36, 7086–7096.
23. Sanakis, Y., Ioannidis, N., Sioros, G., and Petrouleas, V. (2001) *J. Am. Chem. Soc.* 123, 10766–10767.
24. Berthold, D. A., Babcock, G. T., and Yocum, C. F. (1981) *FEBS Lett.* 134, 231–234.
25. Ford R. C., and Evans, M. C. W. (1983) *FEBS Lett.* 160, 159–164.
26. Vass, I., Deak, Z., and Hideg, E. (1990) *Biochim. Biophys. Acta* 1017, 63–69.
27. Buser, C., and Brudvig, G. W. (1992) *Research in Photosynthesis* (Murata, N., Ed.) Vol II pp85–88, Kluwer Academic Publishers, Dordrecht, The Netherlands.
28. Nugent, J. H. A., Demetriou, C., and Lockett, C. (1987) *Biochim. Biophys. Acta* 894, 534–542.
29. Diner, B. A. (2000) *Biochim. Biophys. Acta* 1503, 147–163.
30. Styring, S. A., and Rutherford, A. W. (1987) *Biochemistry* 26, 2401–2405.
31. Kawamori, A., Satoh, J., Inui, T., and Satoh, K. (1987) *FEBS Lett.* 217, 134–138.
32. Pace, R. J., Smith, P., Bramley, R., Stehlik, D. (1991) *Biochim. Biophys. Acta* 1058, 161–170.
33. Lavergne, J., and Etienne, A. L. (1980) *Biochim. Biophys. Acta* 593, 136–148.
34. Sane, P. V., Rutherford, A. W. (1986) *Light Emission by Plants and Bacteria* (Govindjee, Ames, J., and Fork, D. C. Eds.) pp 329–360, Academic Press Inc, New York.
35. Demeter, S., Goussias, Ch., Bernat, G., Kovacs, L., Petrouleas, V. (1993) *FEBS Lett.* 336, 352–356.
36. Kanazawa, A., Kramer, D., Crofts, A. (1992) in *Research in Photosynthesis Vol II* (Murata, N., Ed.) pp 131–134, Kluwer Academic Publishers, Dordrecht, The Netherlands.
37. Ioannidis, N., Sanakis, Y., Sioros, G., and Petrouleas, V. (2001) in *Proceedings of the 12th International Congress on Photosynthesis*, CSIRO Publishing, Collingwood, Australia (www.publish.csiro.au/ps2001; ISBN 0643 067116).
38. Petrouleas, V., and Diner, B. A. (1987) *Biochim. Biophys. Acta* 893, 126–137.
39. Geijer, P., Deak, Z., and Styring, S. (2000) *Biochemistry* 39, 6763–6772.

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